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UltraScript 2.0 Reverse Transcriptase

www.pcrbio.com

Product description:

UltraScript 2.0 Reverse Transcriptase (RTase) is a robust and highly thermostable modified MMLV reverse transcriptase engineered for superior cDNA synthesis speed, yield and representation from a wide range of RNA sample types.

UltraScript 2.0 Reverse Transcriptase is provided with an advanced 5x buffer containing enhancers, dNTPs and MgCl₂ designed to give sensitive and efficient cDNA synthesis from a broad range of RNA input amounts. As oligos are not included, users have the flexibility to define their own priming strategy.

UltraScript 2.0 Reverse Transcriptase is not inhibited by ribosomal and transfer RNAs, making total RNA an ideal substrate. The RTase can be used with 20pg to 3.5µg total RNA or oligo(dT) purified mRNA, however, the optimal template concentration will ultimately be determined by what oligos are used.

The RTase is blended with an advanced RNase inhibitor preventing degradation of RNA by contaminating RNase.

Component	10 000 units	40 000 units
5x UltraScript Buffer	1 x 200µL	4 x 200µL
UltraScript 2.0 (200units/µL) with RNase inhibitor	2 x 25µL	2 x 100µL

Shipping and storage

On arrival the kit should be stored between -30°C and -15°C. Avoid prolonged exposure to light. The kit can go through 30 freeze/thaw cycles with no loss of activity. Avoid exposure of the stock solution to frequent temperature changes and limit handling at room temperature to the necessary minimum.

Limitations of product use

The product may be used only for in vitro research purposes.

Technical support

Help is available on our website at <https://pcrbio.com/resources/> including answers to frequently asked technical questions. For technical support and troubleshooting please email technical@pcrbio.com with the following information:

- Reaction setup
- PCR cycling conditions
- Screen grabs of gel images/real-time PCR traces

Important considerations

5x UltraScript Buffer: Contains 15mM MgCl₂, 5mM dNTPs, enhancers and stabilizers. It is not recommended to add further enhancers or MgCl₂ to the reaction. The buffer composition has been optimised to generate high yield cDNA for downstream applications.

Primers: Suggested primer concentrations are in the table below. For non-biased, non-specific amplification, we recommend using both random hexamers and oligo(dT)₁₈.

Oligo Type	Reaction Concentration	10x Stock Concentration
Specific Primers	1pM	10pM
Random Hexamers	1-5μM	10-50μM
Oligo(dT) ₁₈	50-500nM	0.5-5μM

Template: Use 20pg to 3.5μg total RNA or oligo(dT) purified mRNA for accurate quantification. Additional RNA is not recommended for quantification, as total reverse transcription is not guaranteed. As concentrations of target sequences will vary, users are encouraged to perform a template titration to find the optimal concentration for their application.

Optional preincubation: Incubating template with primers prior to reverse transcription can increase the amount of cDNA, however this step is not necessary for accurate quantification. If preincubation is desired, incubate template with primers for 2 minutes at 70°C, then rapidly cool to 4°C, before adding to reaction.

Incubation temperature: We recommend incubating with a temperature of 50°C for 30 minutes for most applications. Where regions of interest contain high secondary structure (>65% GC), incubation temperatures of up to 70°C may be used, but this will reduce the activity of the enzyme and may result in less total cDNA. The same temperature should be used when comparing samples.

PCR setup: We recommend 4.0μL of cDNA per 20μL PCR reaction. As excess RTase can inhibit Taq activity, better sensitivity can sometimes be obtained by diluting the resulting cDNA. We recommend diluting the cDNA 10x-100x when quantifying genes with low expression.

Reaction Setup

1. Allow 5x UltraScript Buffer to thaw, briefly vortex.
2. Prepare a master mix based on the following table. Insert reagents in the sequence listed:

Reagent	20μL reaction	Final concentration	Notes
5x UltraScript Buffer	4.0μL	1x	
UltraScript 2.0 (200units/μL) with RNase inhibitor	1.0μL		Add before total RNA as RNase inhibitor is blended with RTase
20pg to 3.5μg Total RNA or oligo(dT) purified mRNA	XμL		
10x Primer Mix	2μL	1x	See Primers section
PCR grade dH ₂ O	Up to 20μL final volume		

No RT control setup (optional)

Reagent	20μL reaction	Final concentration	Notes
5x UltraScript Buffer	4.0μL	1x	
20pg to 3.5μg Total RNA or oligo(dT) purified mRNA	XμL		Use equal amount as in step 2
10x Primer Mix	2μL	1x	Use equal amount as in step 2
PCR grade dH ₂ O	Up to 20μL final volume		

Incubation and enzyme denaturation

3. Incubate at 50-55°C for 10-30 minutes.
4. Incubate at 95°C for 10 minutes to denature RTase.